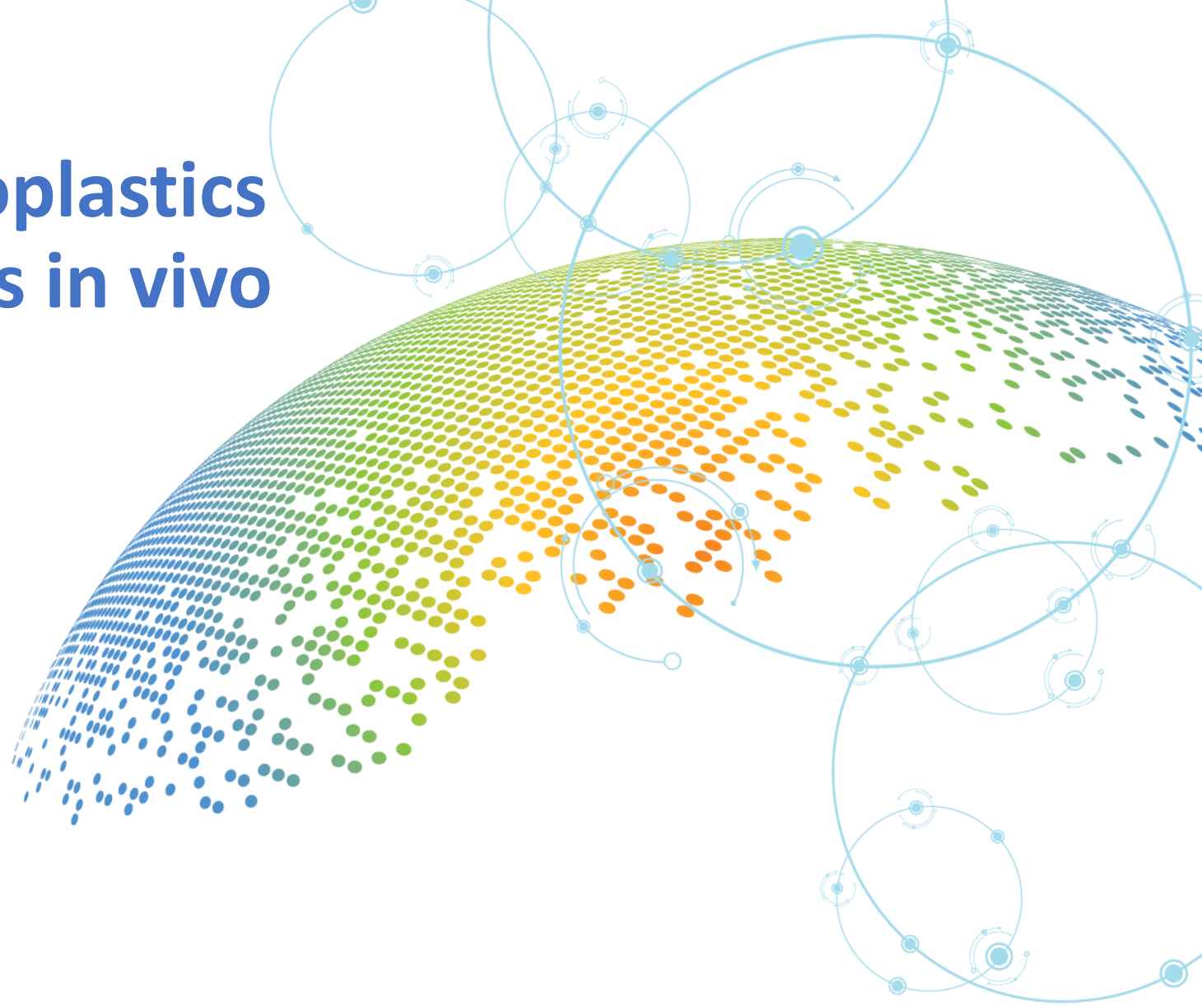


# Well-Characterized Nanoplastics for Oral Exposure Studies in vivo

Leah Johnson, PhD  
RTI International  
Research Triangle Park, NC  
leahjohnson@rti.org



# Outline: Well-Characterized Nanoplastics for Oral Exposure Studies in vivo

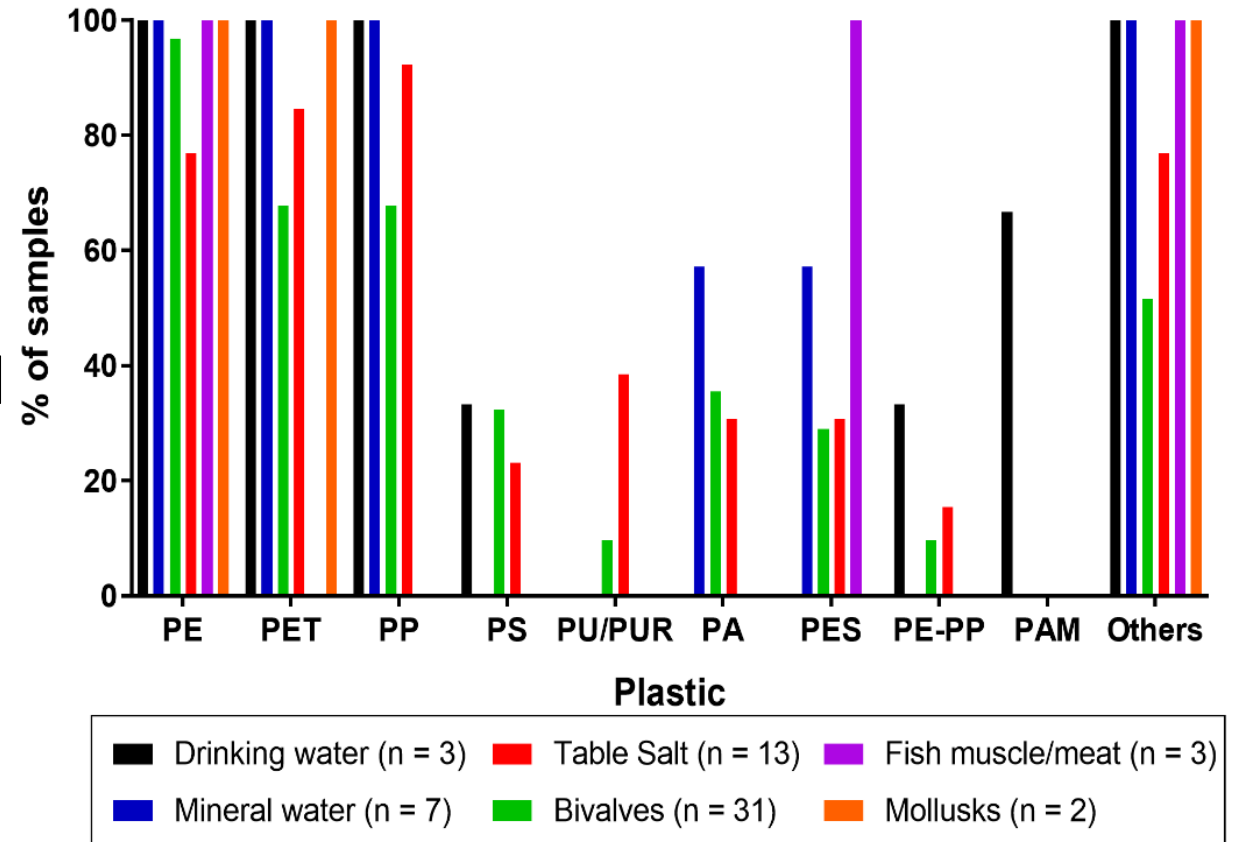
- Brief Background on oral exposure of nanoplastics
- Fabrication/Characterization of in-house fabricated nanoplastics
- Biological Studies at RTI
  - In vitro
  - In vivo oral exposure
- Important Questions/Next Steps



Image from Nat. Geo. exhibit, 'Plant or Plastic' Norfolk, VA.

# What Plastics Are We Exposed To?

- Review of small-scale plastics detected in drinking water, Beverages, Food Sources (<math><50 \mu\text{m}</math>)
- Mixtures in samples
  - >35 types of polymers reported in drinking water, beverages, and food sources.
  - Common polymers reported include polyethylene (PE), polypropylene (PP), polyethylene terephthalate (PET), polyethersulfone (PES), polyamide (PA), polystyrene (PS).



Mortensen NP, TR Fennell, LM Johnson (2021) NanoImpact.

# Oral Exposure of Polystyrene (PS) Particles & Gastrointestinal Absorption in Rats

## Jani et al '89

Fluorescent PS particles; oral gavage; 100nm – through 1  $\mu$ m detected in Peyer's patches, villi, liver, lymph nodes, spleen.<sup>1</sup>

## Jani et al '90

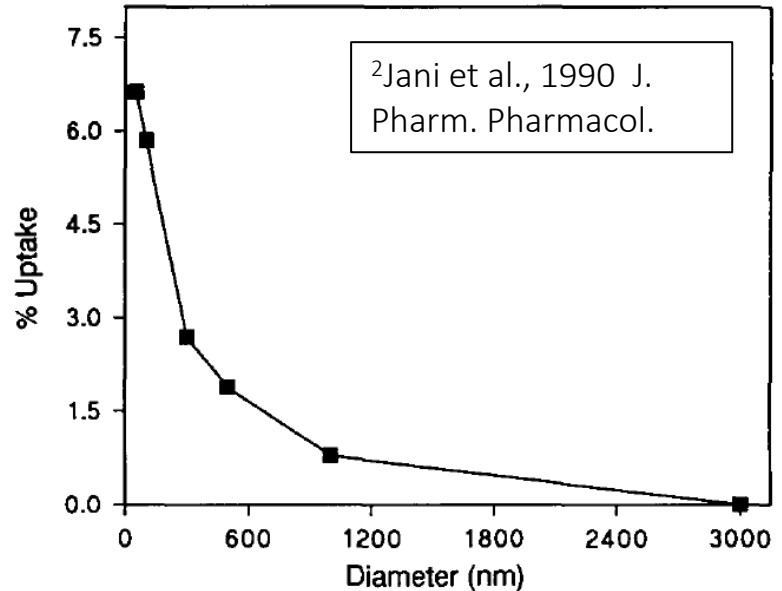
Quantify oral absorption/location of PS different sizes; increases with decreasing size for PS spheres (50 nm - 3  $\mu$ m), in rats.<sup>2</sup>

## Jani et al '96

Biliary elimination of FITC labeled PS spheres (50, 500 nm, 1, 3  $\mu$ m); size-related excretion of PS spheres into the bile.<sup>3</sup>

Translocation of PS across gastrointestinal wall and uptake of PS (<50 nm) within systemic organs.

Cumulative uptake in liver, spleen, blood, bone marrow and kidney



<sup>1</sup>Jani et al. 1989 J. Pharm. Pharmacol. DOI: 10.1111/j.2042-7158.1989.tb06377.x

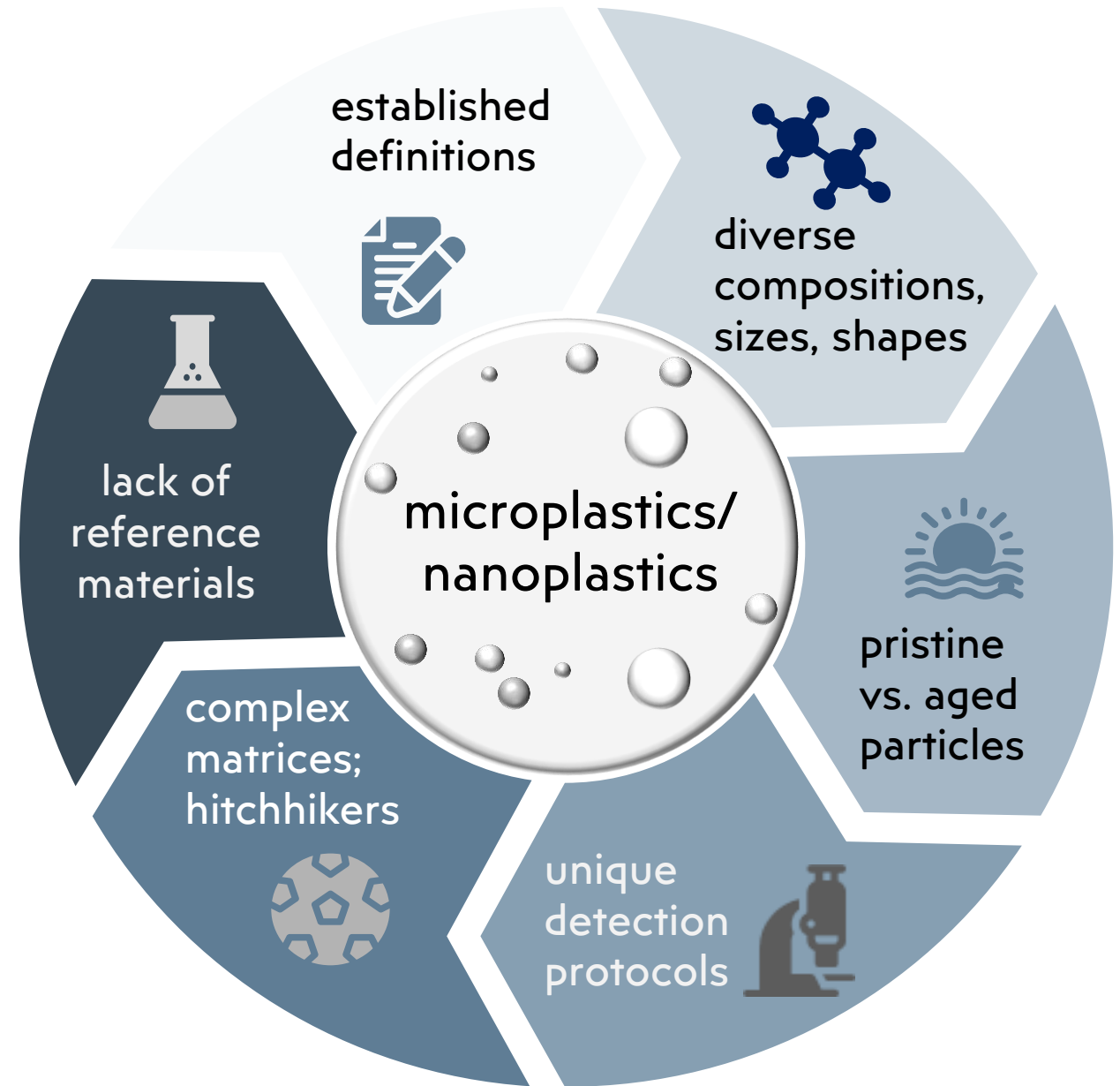
<sup>2</sup>Jani et al., 1990 J. Pharm. Pharmacol. DOI: 10.1111/j.2042-7158.1990.tb07033.x

<sup>3</sup>Jani et al., 1996 J. Drug Targeting. DOI: 10.3109/10611869609046266

# Factors to Consider for Micro/Nanoplastics Research

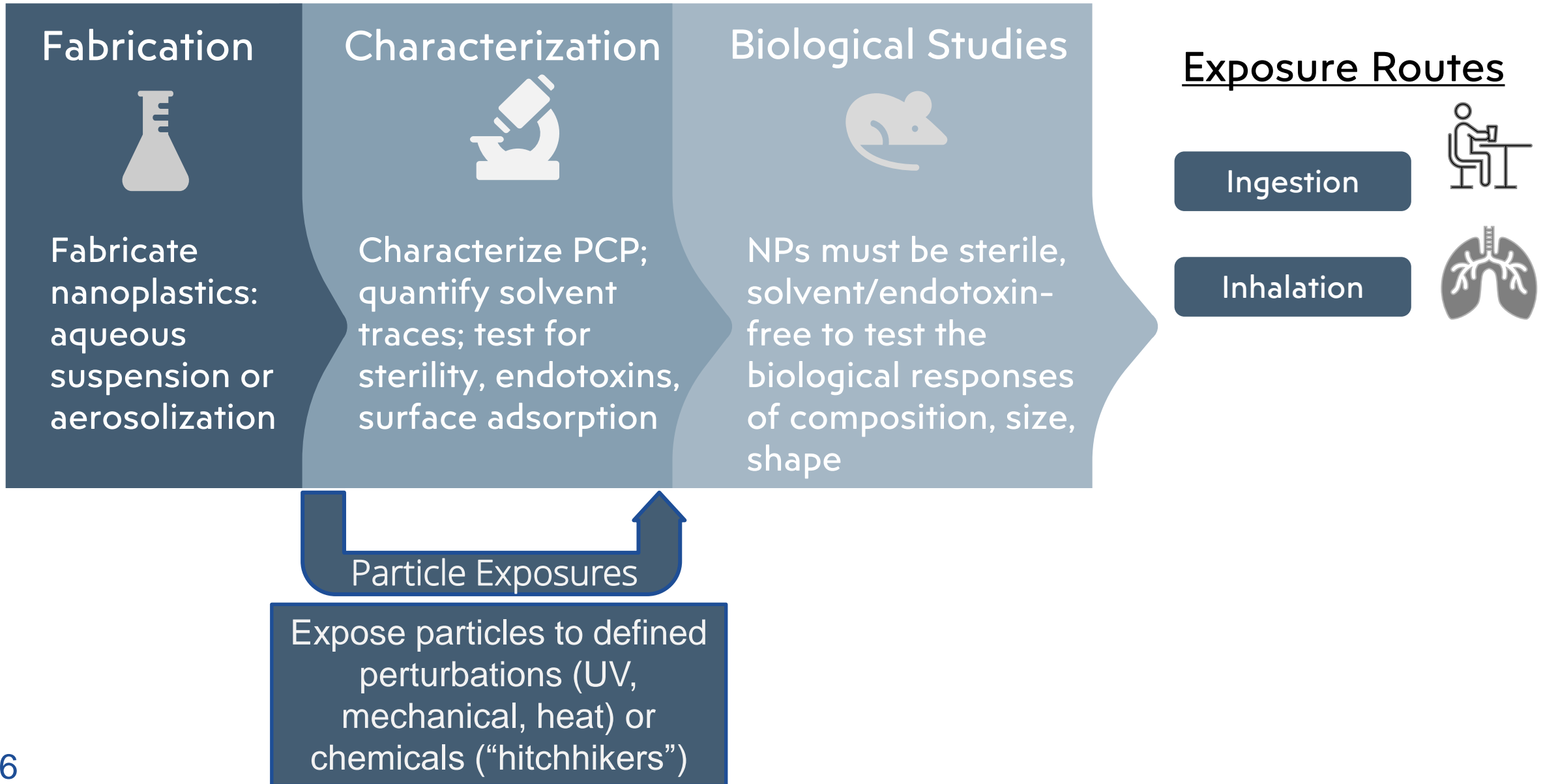
*Diversity & unique attributes of microplastics & nanoplastics require new approaches for conducting studies*

How do we deal with these variables?



Adopted from Paul, Maxi B. et al. *Nanoscale Advances* 2 (2020): 4350.

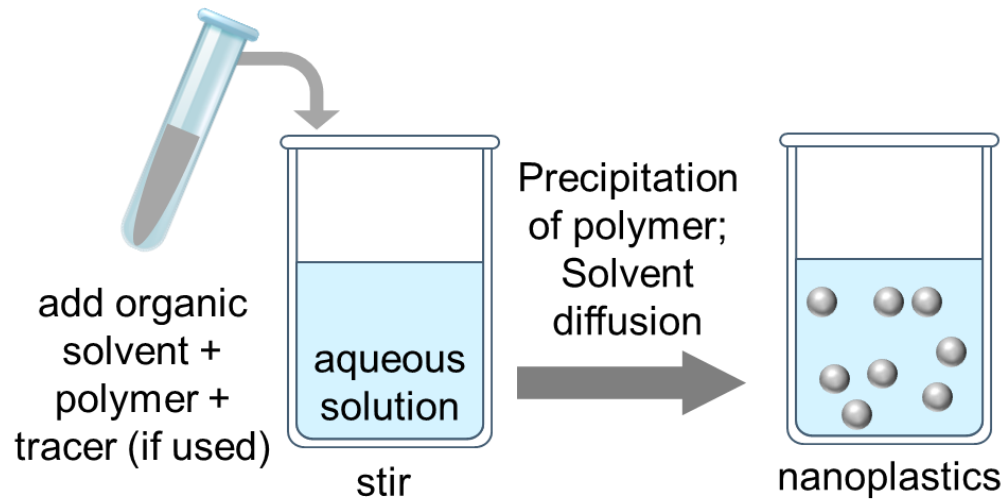
# Approach: Systematic Testing with Well-Characterized NPs



# Fabrication of Nanoplastics

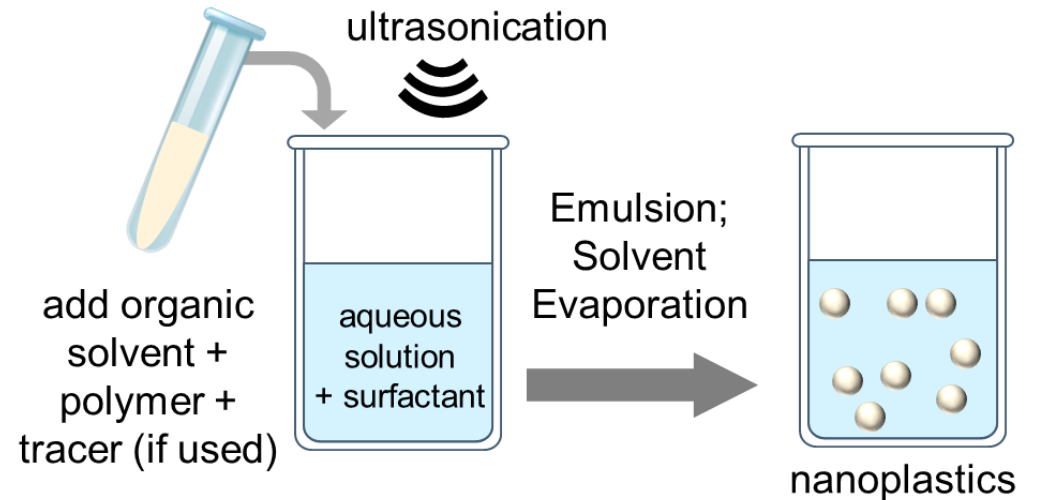
## General Approach: Dispersion of Preformed Polymers

### Nanoprecipitation



**Advantages:** rapid, single-step, low-energy, reproducible, amenable to scale-up

### Emulsion/Solvent Displacement



**Advantages:** rapid and useful for more hydrophilic polymers, but requires a high shear

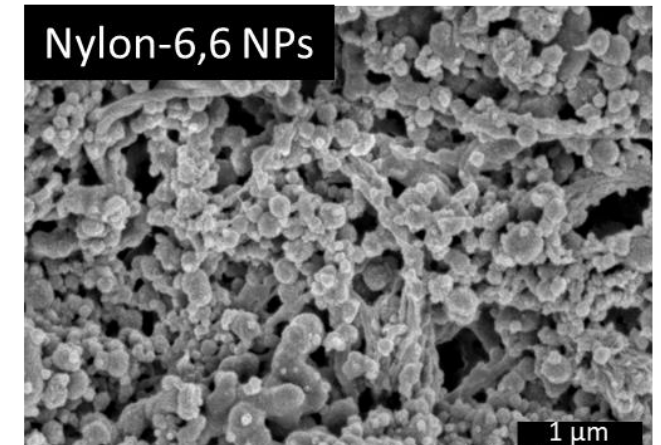
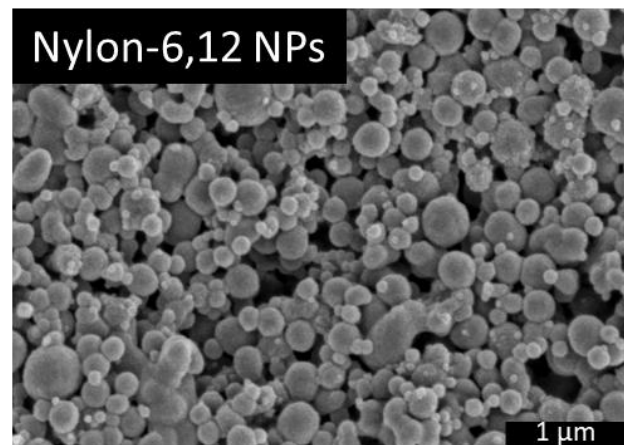
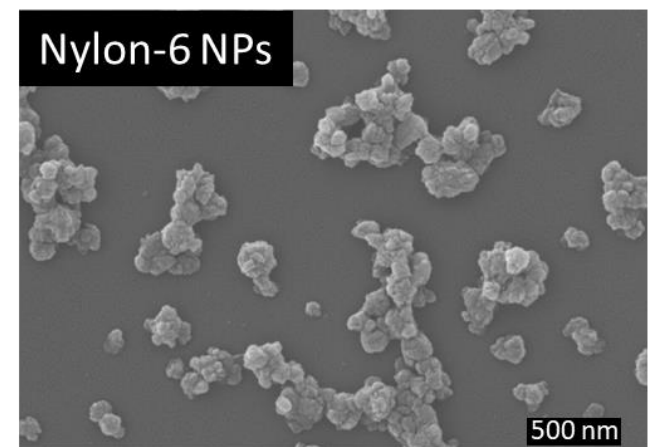
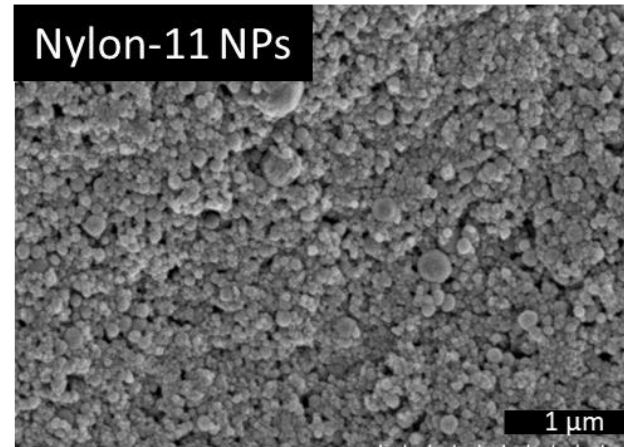
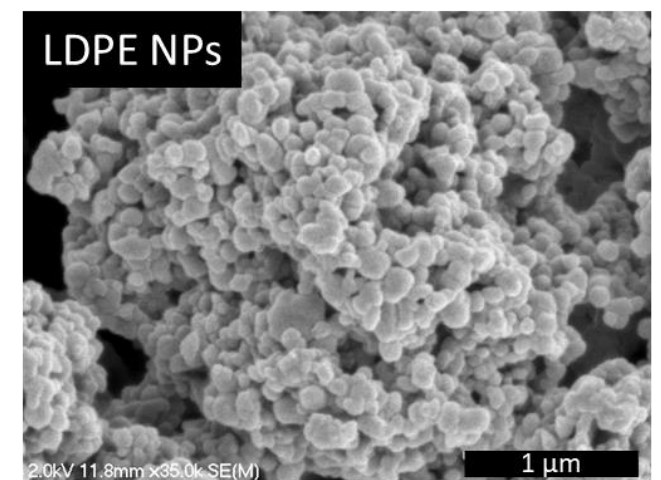
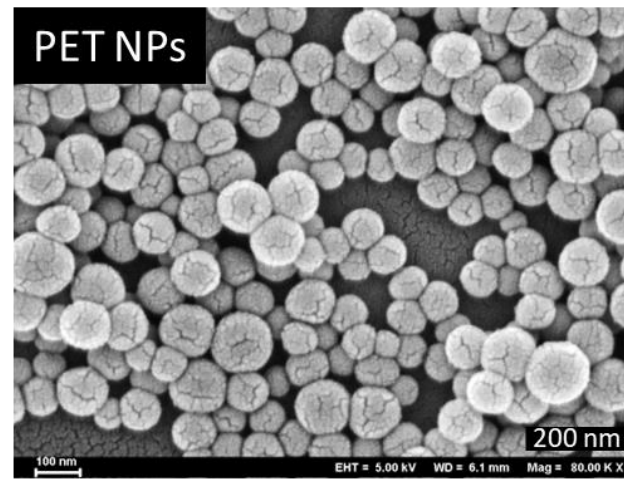




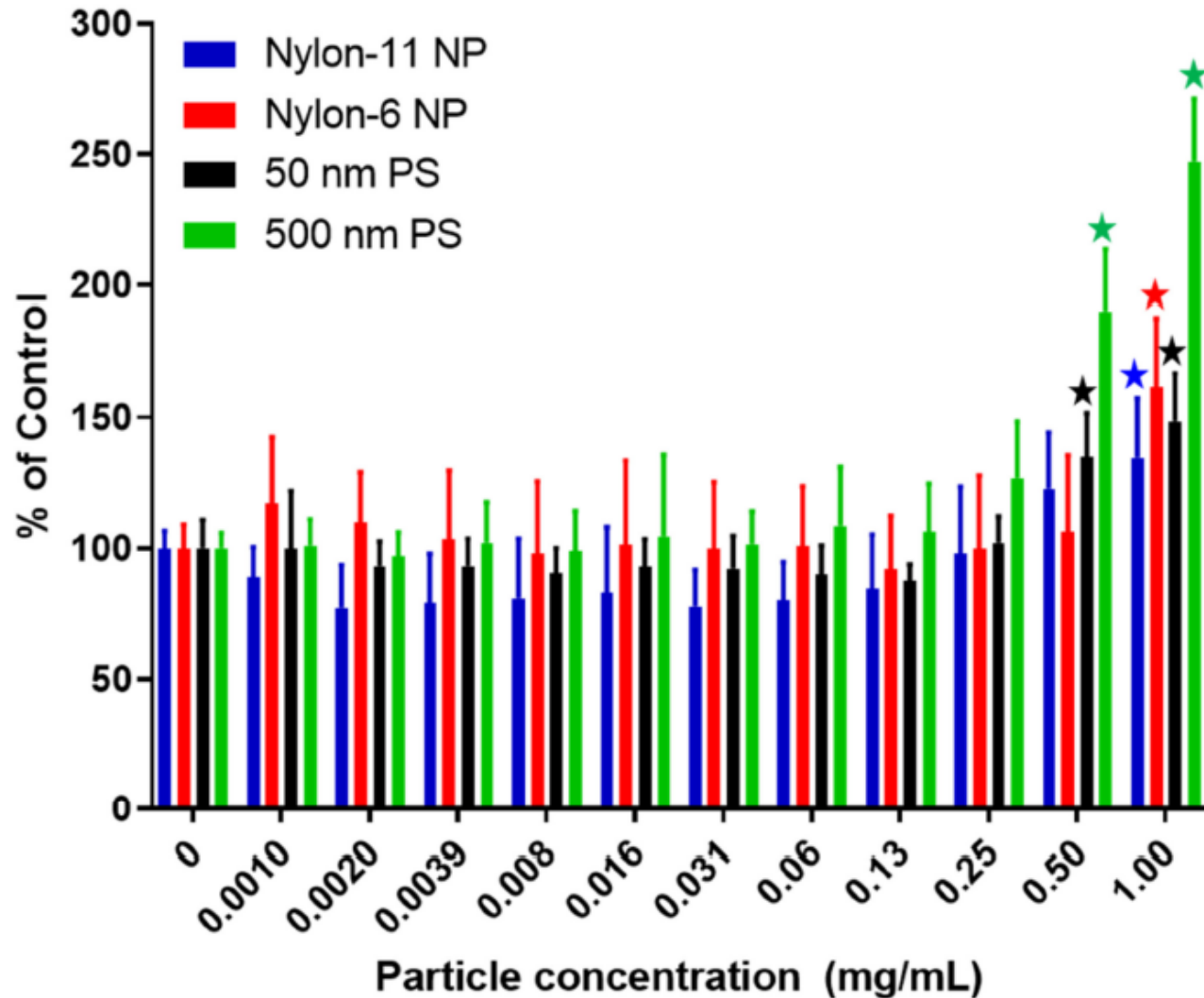
# Characterization of NPs

Thorough characterization of nanoplastics prior to initiating testing in biological systems

- ✓ **Microscopy-** morphology, fluorescence
- ✓ **FT-IR-**characteristic absorption bands of bulk polymer
- ✓ **GC/MS:** characteristic peaks
- ✓ **DLS:** hydrodynamic diameters
- ✓ **Zeta Potential** in various solutions
- ✓ **Endotoxin:** > 0.5 EU/mL
- ✓ **Leaching studies:** ensure stability of tracer incorporation.



# Nylon NPs in RAW264.7 Macrophages



## Polyamide (nylon) Properties

- Diverse chemical family
- Susceptible to strong acid/alkaline
- High toughness

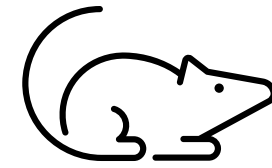
## Products

- Films for Food Packaging
- Filaments (toothbrush bristles)
- Cosmetics
- Tubing for beverages
- Fabric and Fibers (clothing, flooring)

## Cytotoxicity Assay

- Cytotox. via cell membrane integrity (LDH)
- PS NPs: no significant cytotoxic effects up to 0.25 mg/mL
- Nylon NPs: no cytotoxicity until high doses (1mg/mL)
- 500 nm PS at (1 mg/mL) interfered with LDH assay

# In vivo Evaluation of NPs in Sprague Dawley Rats



**Purpose:** Investigate potential biological effects of NPs (nylon, PS) following oral exposure early in life

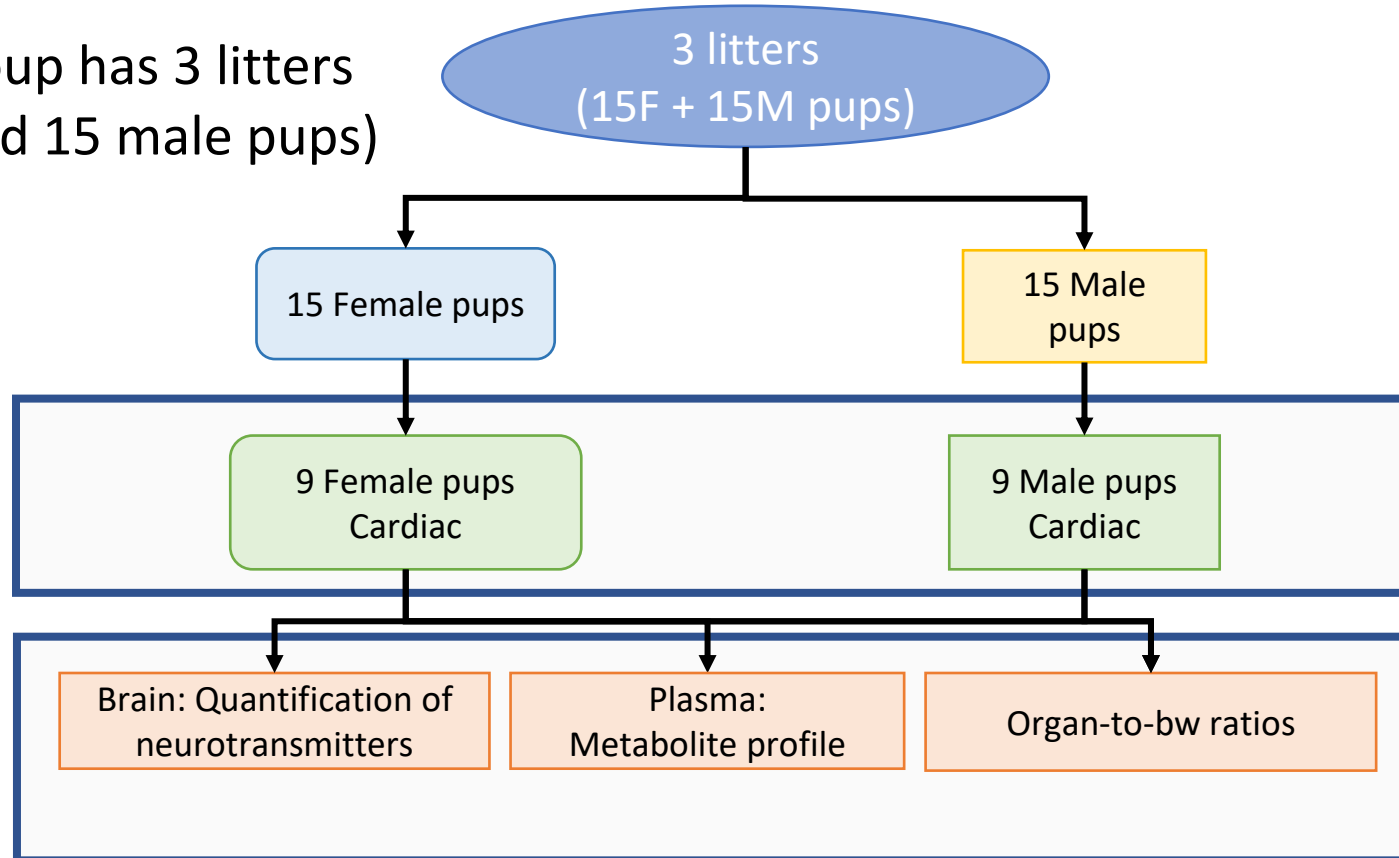
## Study design:

Each dose group has 3 litters (15 female and 15 male pups)

PND 7 – 10:  
Gavage daily (4 doses)

PND 20:  
Non-invasive assay

PND 21:  
Necropsy



## Dose Groups:

- Vehicle control (H<sub>2</sub>O)
- 20 mg/kg bw/day Nylon-11 NP
- 20 mg/kg bw/day PS NP

## Nanoplastic characterization:

- Endotoxin
- Size and surface chemistry
- Composition

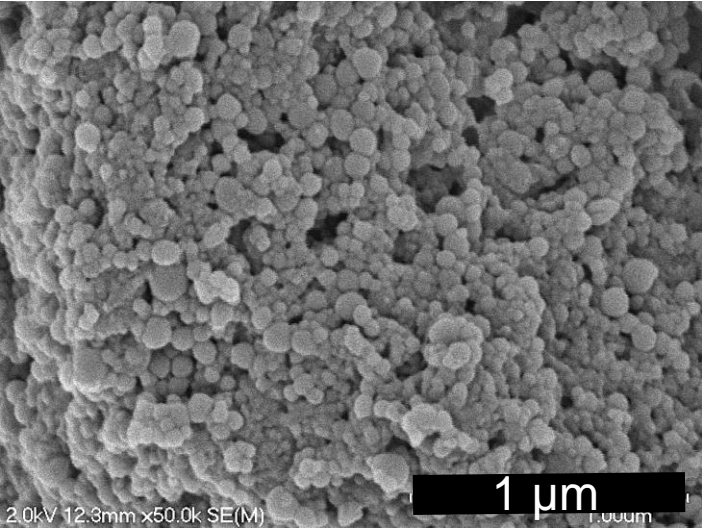
## Analysis:

- Body weight (bw)
- Cardiac assessment: ECG
- Organ-to-bw ratio (brain, liver)
- Brain: Neurotransmitters
- Plasma: 186 metabolites

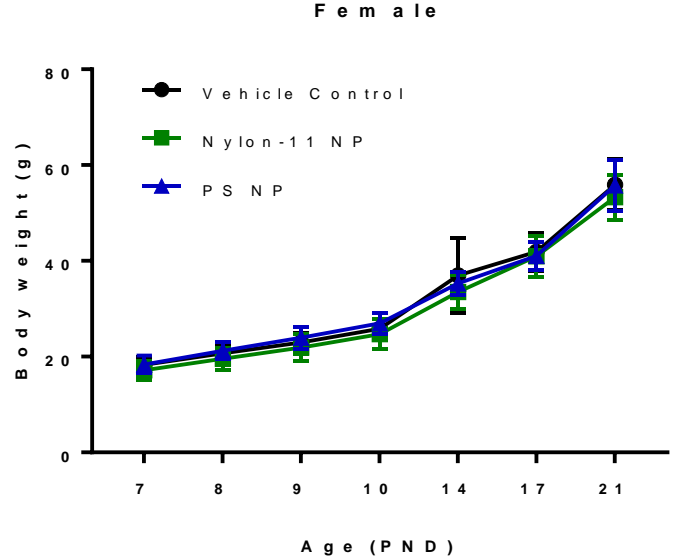
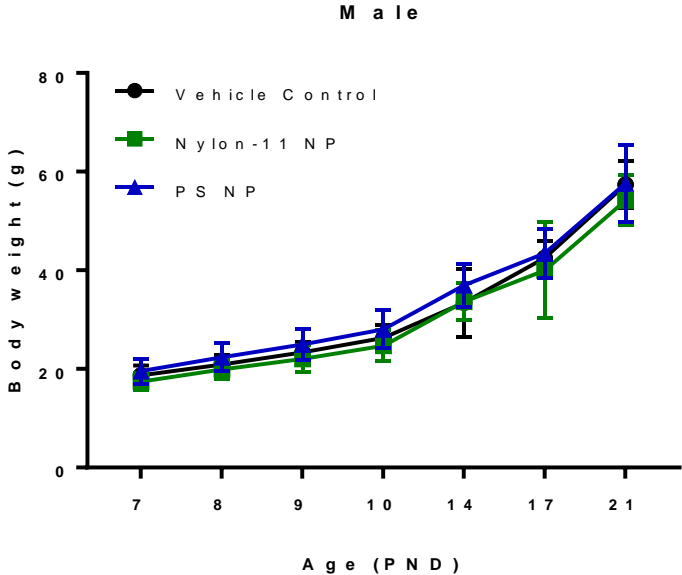
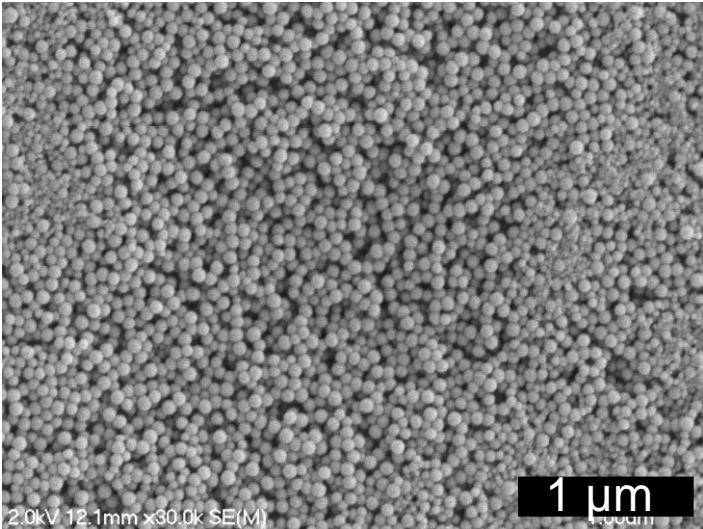
# In vivo Evaluation of NPs in Sprague Dawley Rats: Preliminary Results



**Nylon-11 (RTI)**



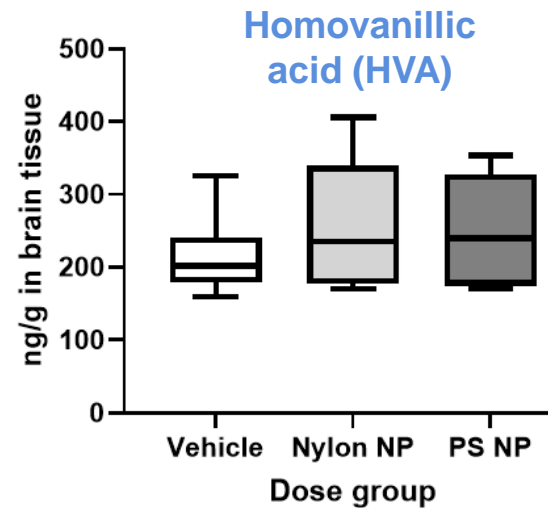
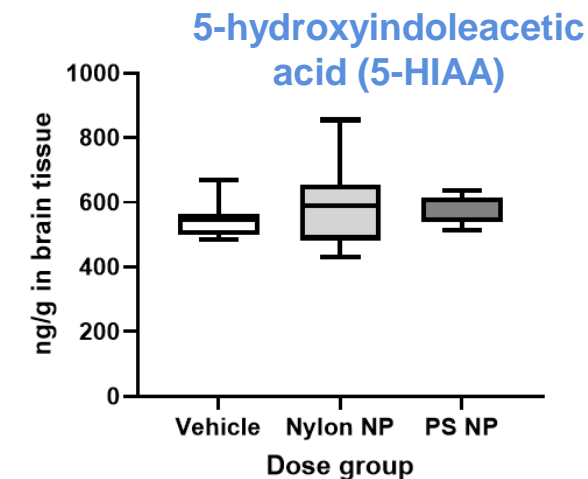
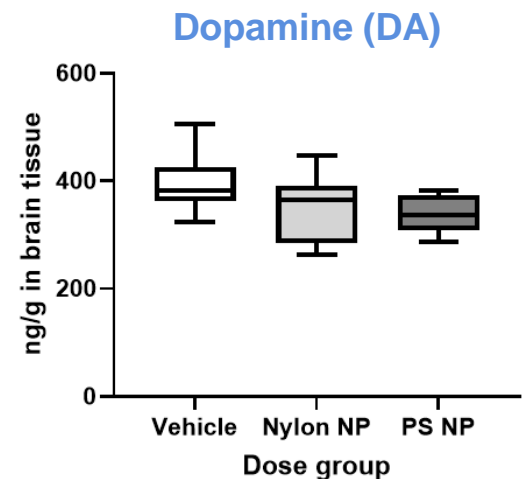
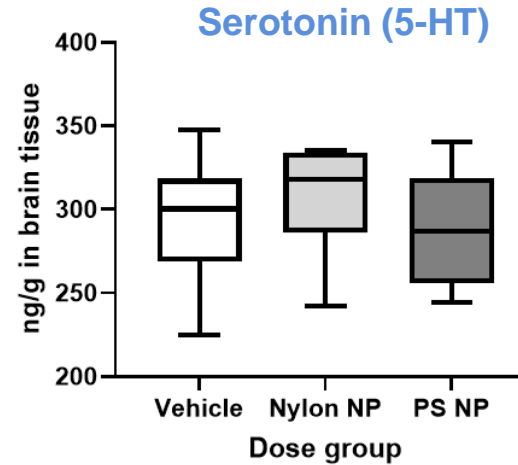
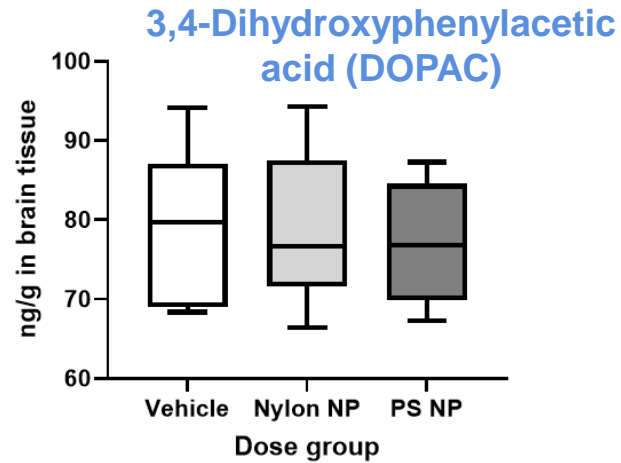
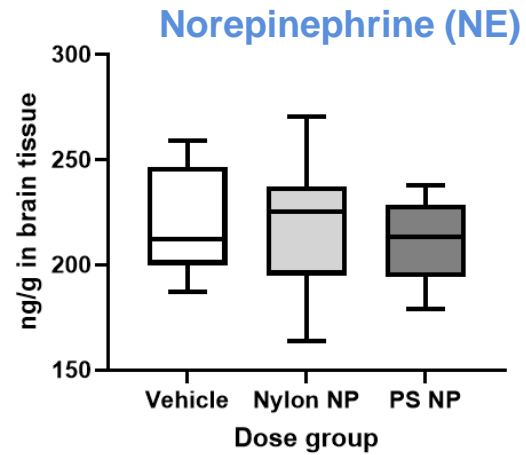
**Polystyrene (Spherotech)**



NP	Average size (nm)	Average PDI	Average zeta potential (mV)	Endotoxin (EU/mL)*
Nylon-11	139 ± 44	0.2 ± 0.04	28 ± 0.6	0.054
Polystyrene (PS)	96 ± 21	0.04 ± 0.02	-49 ± 0.3	0.048

*Endotoxin values below the recommended FDA limit of 0.5 EU per mL for medical devices*

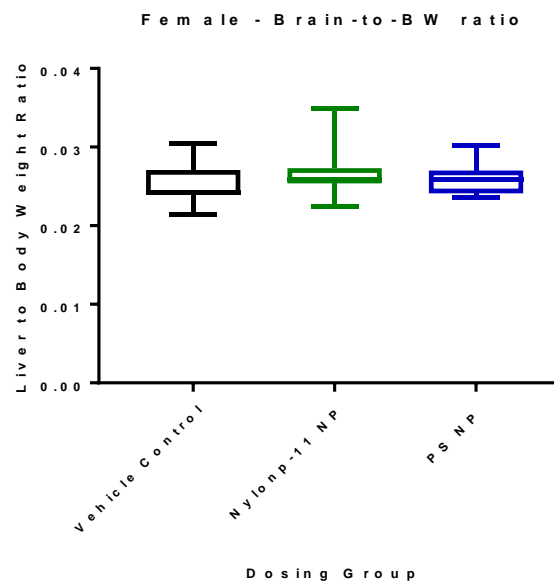
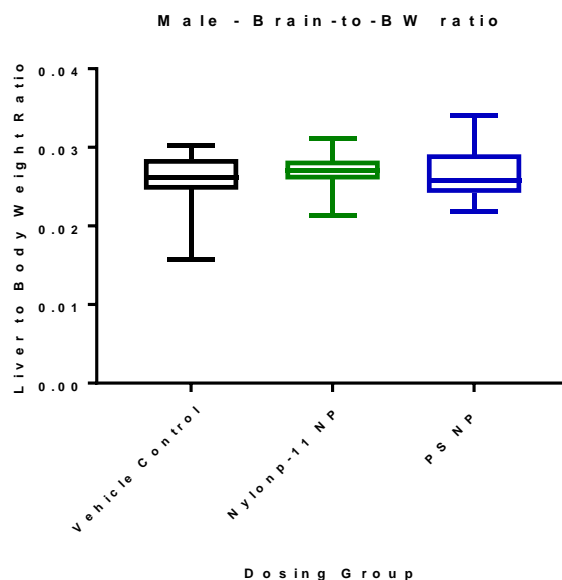
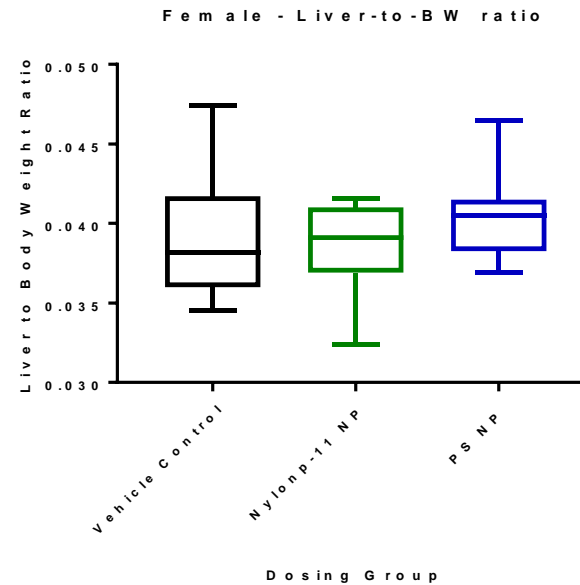
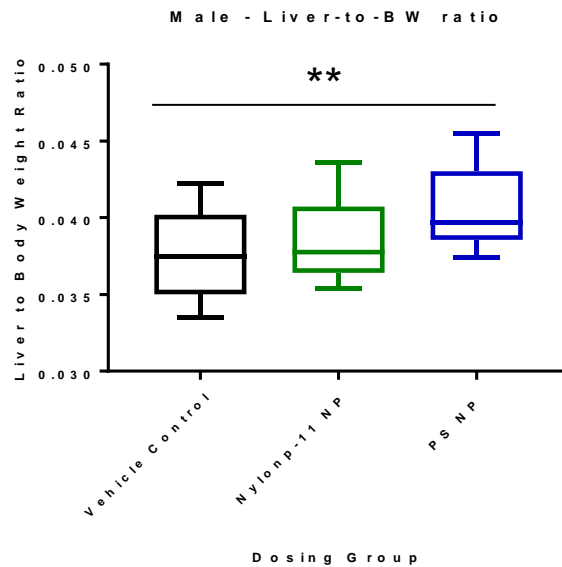
# Neurotransmitter Concentration in Brain: Female Sprague Dawley Rats



## Preliminary Results

- Concentration of 6 monoamine neurotransmitters/metabolites determined in right-brain for male and female pups at PND 21
- No significant changes were found in either sex for any analytes.

# In vivo Evaluation of NPs in Sprague Dawley Rats

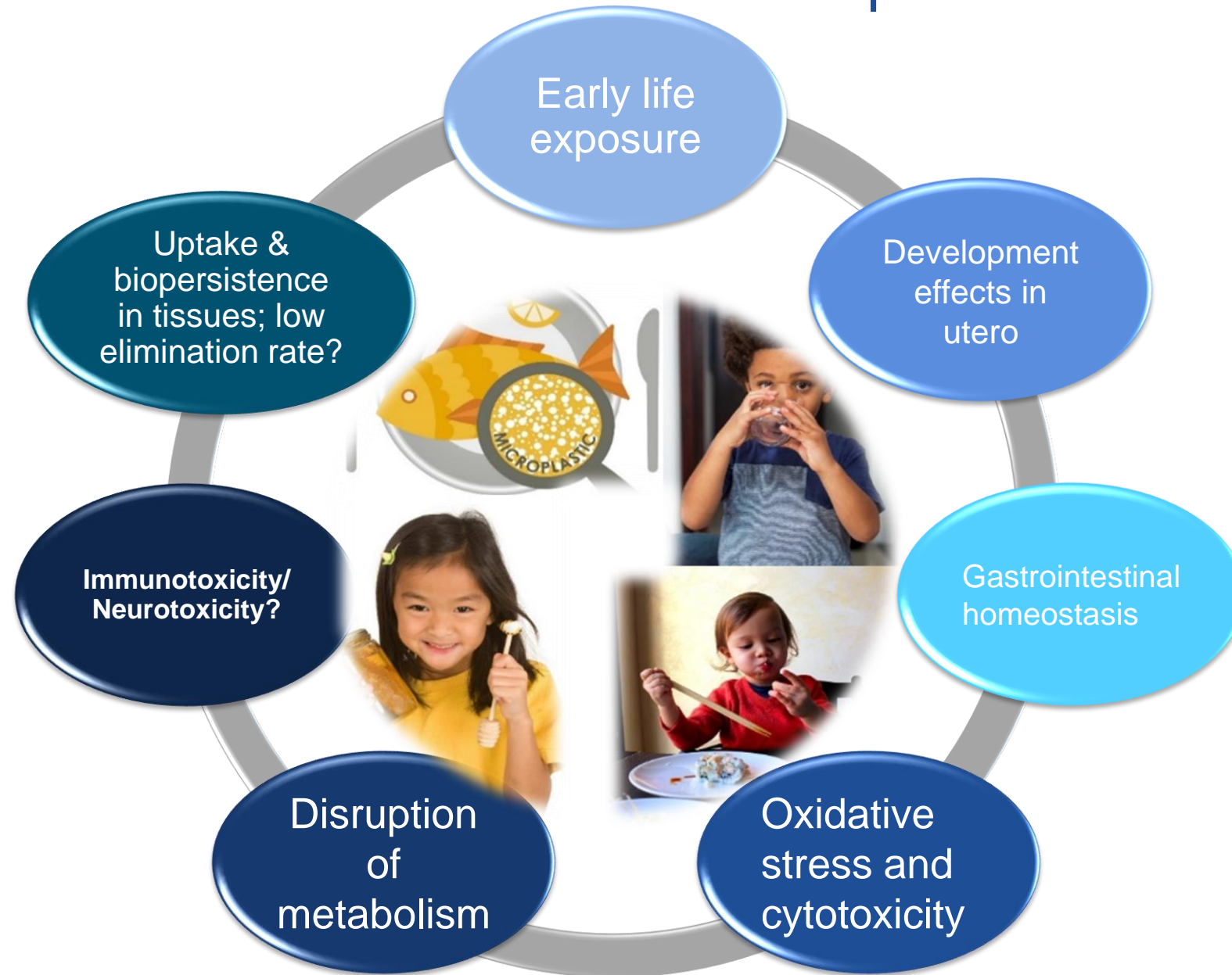


**Liver/BW Ratio:** useful endpoint to detect organ toxicity.  
**Brain/BW Ratio:** Brain undergoes growth/development early in life; multiple events can impact biochem/homeostasis in the brain

## Preliminary Results:

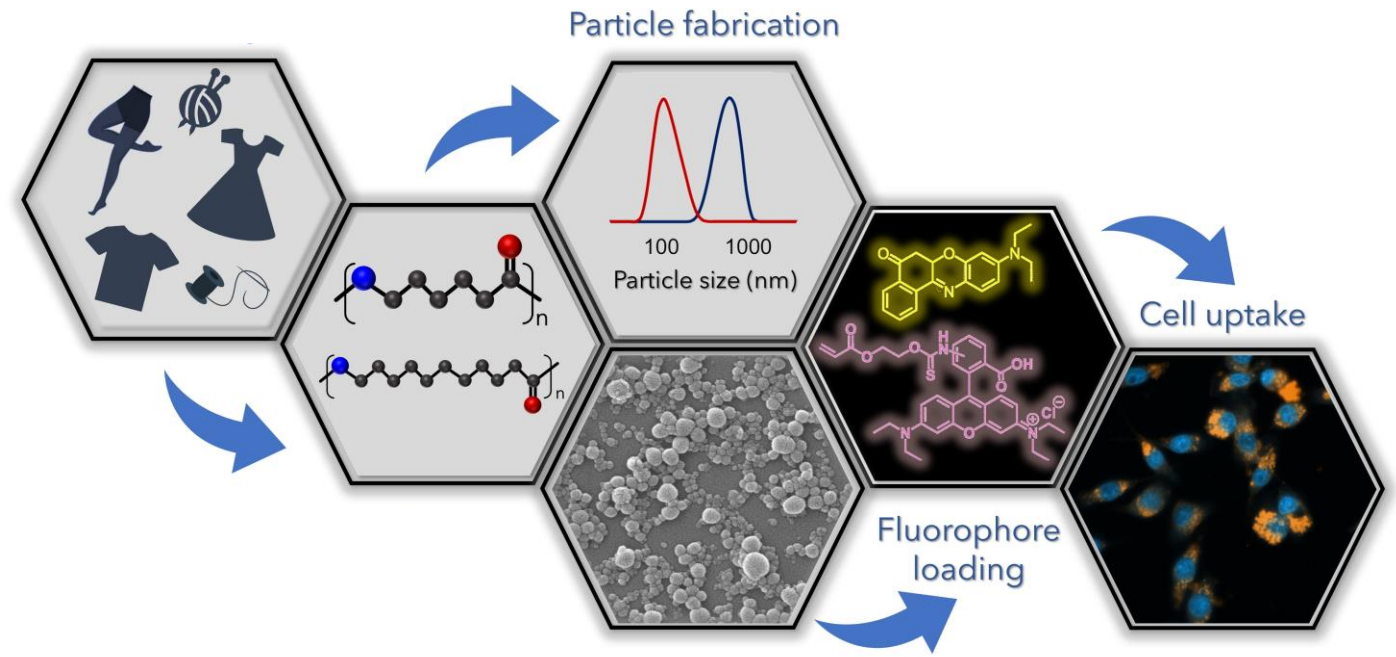
- Increased liver-to-BW ratio for males administered with PS NPs as compared to the vehicle control.
- No differences for other analytes

# Questions: Potential Health Risks of Oral Exposure to NPs



# Future Research Needs

- A better understanding of exposure and effects.
- Reference materials reflecting the plethora of NP and MP identified as sources of human exposure.



- Development of sampling strategies, extraction, characterization, and identification.
- Systematic studies of the role of PCP including size and polymer chemistry plays in uptake and interference with biological functions.
- Prioritization of which NP and MP should be studied.
- Removal of existing NP and MP from the environment and sources of human exposure.



# Acknowledgements and Contributors

## Contact

Leah Johnson  
leahjohnson@rti.org

**Archana Krovi, PhD**  
Senior Research Chemist



**Ninell Mortensen, PhD**  
In Vitro; Exposure



**Timothy Fennell, PhD**  
Senior Director



**Leah Johnson, PhD**  
Senior Director,  
Biomedical Technologies



Funding sources:



**Ryan Chartier, BS**  
Research Chemist



**Jeff Mecham, PhD**  
Research Chemist



**Maria Moreno, BS**  
Chemist



---

## ***Materials Characterization***

---

*Dr. Jean Kim, Dr. Sara Maloney, Ms. Brenda Fletcher,  
Mr. Rodney Snyder, Mr. Randy Ruthowske, Ms. Randi  
Carter, Mr. Alex Kovach, Ms. Sarah Harrison, Dr. Ian  
Stewart, Dr. Shyam Aravamudhan*

---