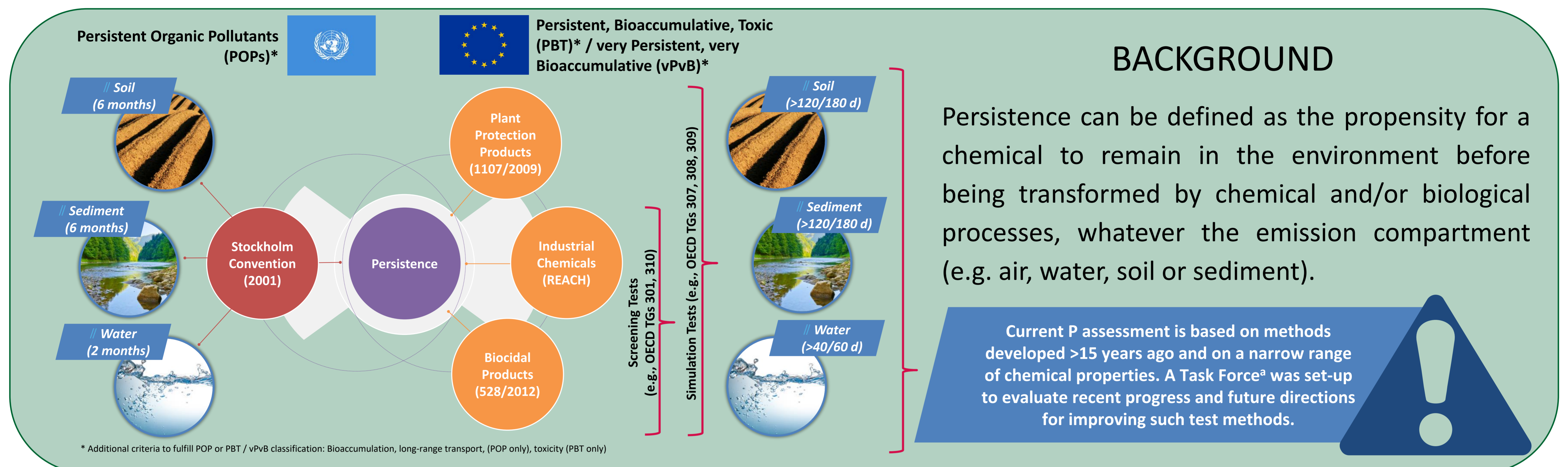


Scientific concepts and methods for moving persistence (P) assessments into the 21st century

Russell Davenport (Newcastle University, UK); Pippa Curtis-Jackson (Environment Agency, UK); Philipp Dalkmann (Bayer, DE); Jordan Davies (LyondellBasell, NL); Kathrin Fenner (EAWAG, CH); Laurence Hand (Syngenta, UK); Philipp Mayer (Danish Technical University, DK); Kathleen McDonough (Procter & Gamble, US); Andreas Schäffer (RWTH Aachen University, DE); Cyril Sweetlove (L'Oreal, FR); Amelie Ott (Newcastle University, UK); Jens Bietz (Clariant, DE); John Davis (Dow, US); Delina Lyon (Shell, US); Jens Otte (BASF, DE); Frédéric Palais (Solvay, FR); John Parsons (University of Amsterdam, NL); Neil Wang (Total, FR); Johannes Tolls (Henkel, DE); Tim Gant (King's College London, UK); Aaron Redman (ExxonMobil, BE). E-mail: aaron.d.redman@exxonmobil.com



CHALLENGES

Substrate-specific factors

Particles & poorly solubles: Need new testing procedures (problems: sedimentation, sorption, desorption)¹

Non-extractable residues (NER): Methodology for differentiation of the three binding types not yet validated²

UVCBs: Difficult testing of complex substances (ecotox, biodegradability) without knowing each component³



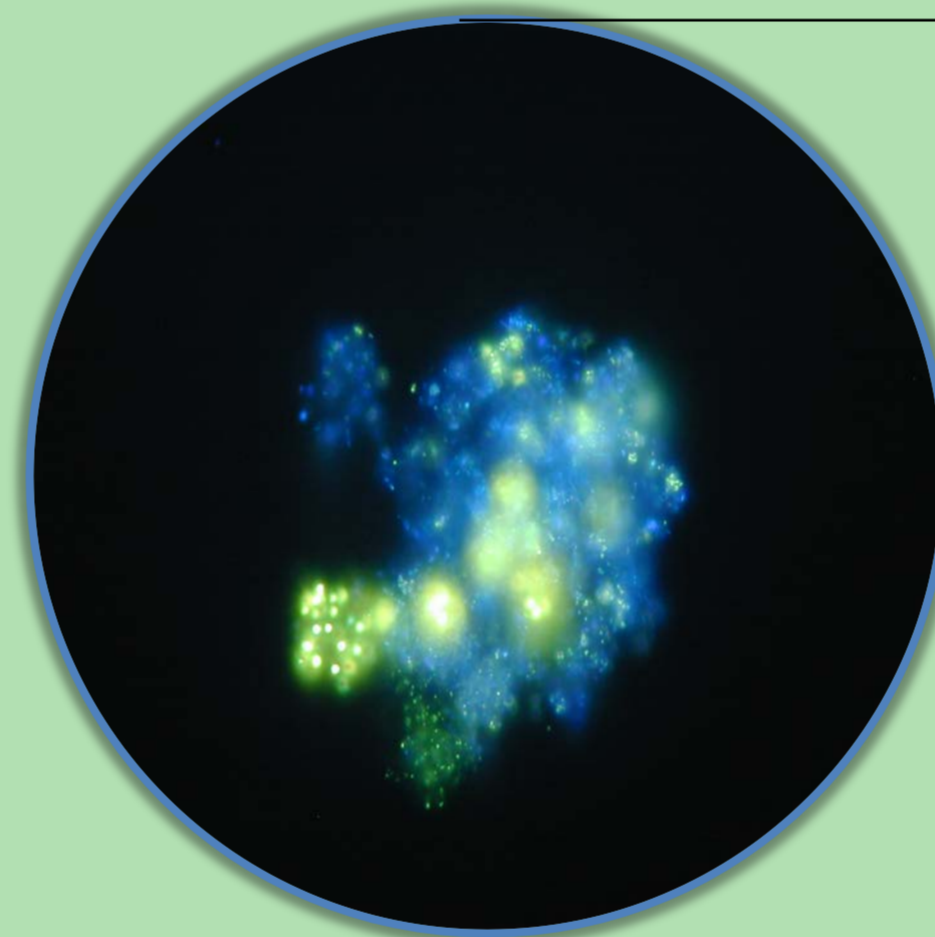
- Poorly solubles: Passive dosing allows realistic and constant exposure concentrations⁹
- NER: Isotope-label studies allow to distinguish primary vs ultimate degradation, types of binding, and establish mass balance and remobilisation potential²
- UVCB: Whole substance ecotox testing, biodegradability testing (based on carbon balance approach), read-across and further *in-silico* tests, 'omics' analyses (targeted and non-targeted)³

Microbial-specific factors

Sample sizes are small (up to 10,000 × less cells) leading to large variability in outcomes⁴ and half-lives⁵ in biodegradation tests, which has an impact on diversity

Diversity, biogeography & lifestyles: Chance inclusion of relatively rare catabolic functions⁴ influenced by environmental compartment/location/redox

Adaptation is currently excluded from biodegradation screening tests⁶



- Increased number of total cells can improve screening test accuracy¹⁰.
- Biomass measurements for normalization of tests¹⁰ and/or degradation rates across test formats¹¹.
- Microbial community analyses ('omics') to contextualize, normalize and understand biodegradation rate variations¹².
- New adaption tests improve the accuracy of screening outcomes¹³

Testing format & abiotic environmental factors

Photolysis is rarely studied⁷ (except for PPPs⁸), but is important for a complete understanding of potential persistence

Temperature: Testing is usually done at a single temperature^{7,8}, but degradation processes are often temperature-dependent (e.g., biotransformation)



- Performance of tests on abiotic processes (e.g., hydrolysis (OECD 111¹⁴), photolysis (OECD 316¹⁵))
- Inclusion of abiotic processes in simulation tests (e.g., irradiation in OECD 309 tests)¹⁶
- Inclusion of abiotic data in weight-of-evidence approach

ADDITIONAL GENERIC TOOLS AND APPROACHES

Benchmarking

Benchmark chemicals for assessing biodegradation relative to a reference chemical(s).¹⁷

Modelling

- High-quality biodegradation rate database to include metadata¹⁸
- Inverse modelling of rates from degradation/dissipation data¹¹
- Further development of QSBRs grouping chemicals by reaction type¹²

Biochemical assays

Screening for specific transformation types: hydrolysis, redox, substitutions with/without enzymes (cf plant metabolism¹⁹)

NEXT STEPS

- ECETOC Task Force^a is on-going and will draft two publications for researchers and practitioners on: i) a conceptual framework for persistence^{Poster 3.17P.7} and ii) introduce more recent improvements in test methods.
- A workshop for academia, industry and regulators is scheduled for September 2020 to further discuss our findings.

^a<http://www.ecetoc.org/taskforce/moving-persistence-p-assessments-into-the-21st-century/>